

# Conifer Translational Genomics Network Coordinated Agricultural Project



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Genomics in Tree Breeding and Forest  
Ecosystem Management

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Module 12 – Marker Informed Breeding  
(MIB) – Program Management



***Nicholas Wheeler– Oregon State University***

# Marker applications

- Quality control
- Introgression and hybrid breeding
- Parentage analysis
  - *Pollen contamination*
- Enhanced breeding designs

# Quality control in a clonal seed orchard

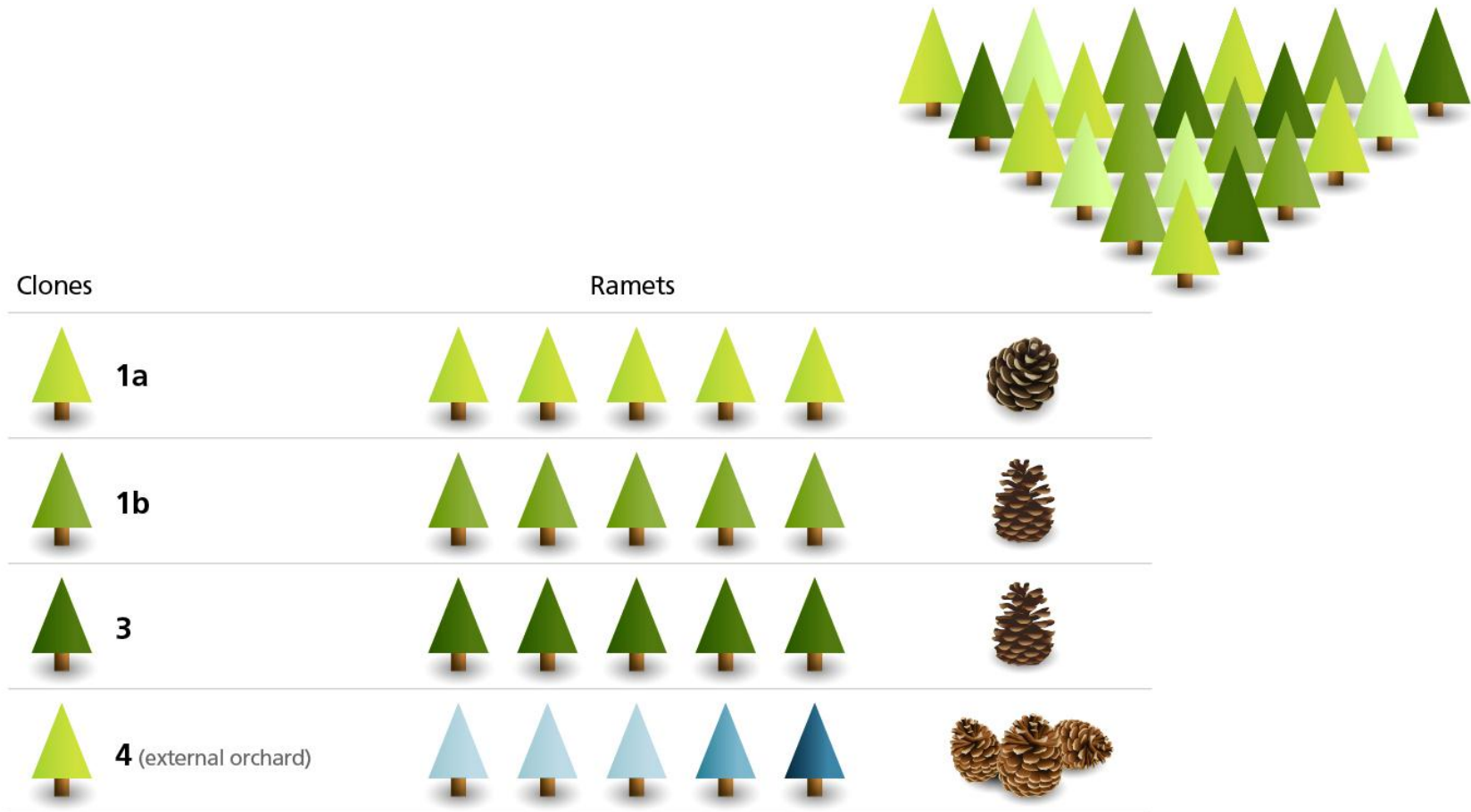


Figure Credit: Nicholas Wheeler, Oregon State University

# Opportunity cost of orchard errors

## Assume

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- Progeny of wrong orchard clone have 5% less genetic gain (volume) than progeny of desired clone
- 1% gain = \$10 present value/acre planted (\$60 future value at rotation)
- Annual planting requirement = 10,000 acres, of which 10% is planted with the desired clone (1000 acres)
- 40% of trees in desired clone are mislabeled; thus, 400 acres per year are planted with trees that are performing 5% below expected
- $400 \text{ acres} * 5\% * \$10/\text{acre/percent} = \$20,000$  present value (\$120K future value). If unchecked for 15 years, = \$300K present value

# Marker assisted backcrossing or introgression

- This is easily the single greatest application of markers to animal and plant breeding in the world
- Used predominantly for simply inherited traits such as disease or insect resistance
- Commonly used to introduce an important allele from an unimproved individual / species into a highly selected individual / line / variety

# Backcross breeding approaches

- Marker-assisted “foreground” selection
  - *Select for markers that identify favorable alleles from the donor population*
  - *Markers must be in very tight linkage with desired trait*
- Marker-assisted “background” selection
  - *Select for marker alleles that identify the host (recurrent) parent genome*
  - *Must have many alleles with complete genome coverage*
- Typically, both approaches are used together

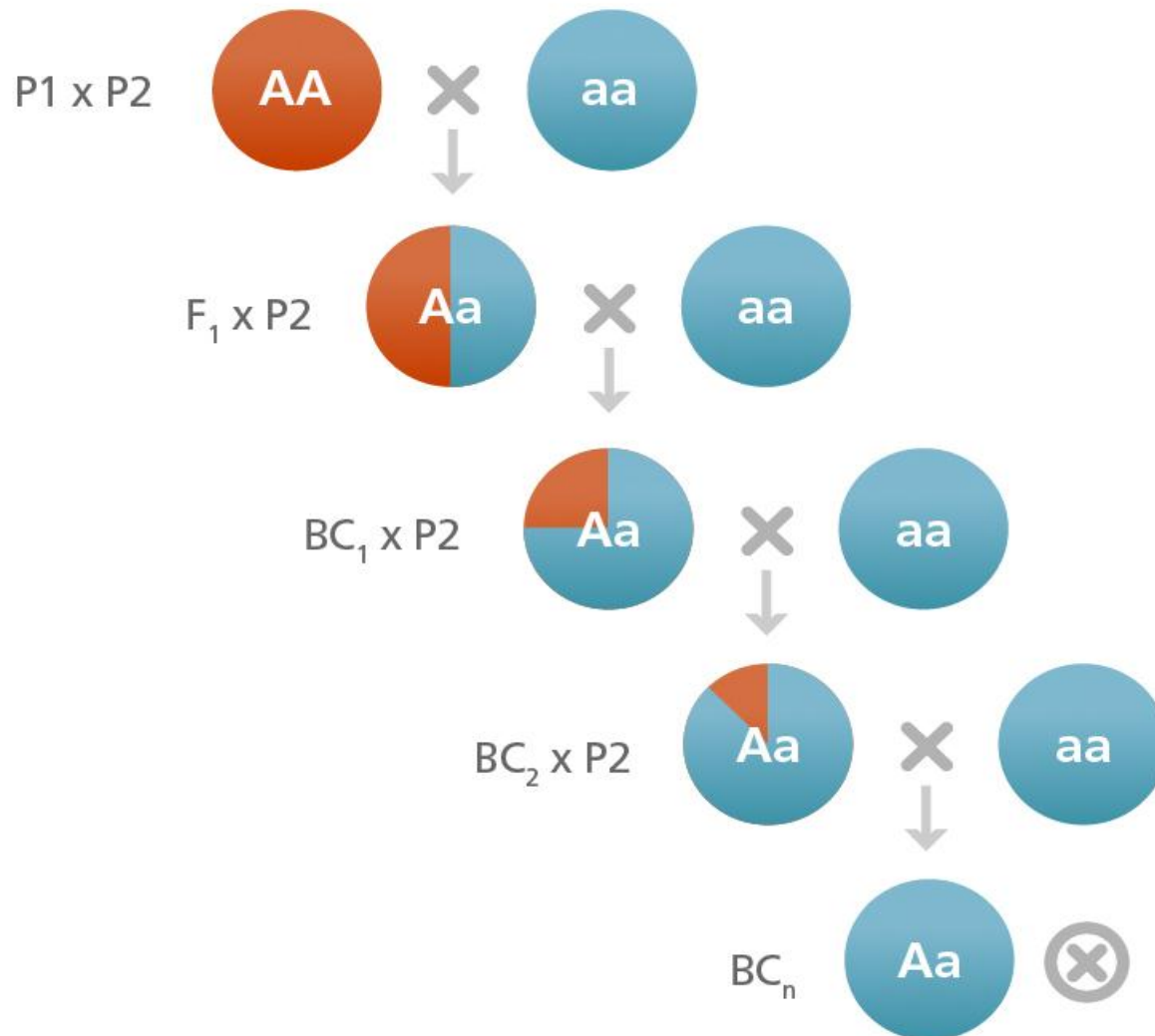


Figure Credit: Modified from Welz and Geiger, 2000

## Distribution of genotypes in simulated BC<sub>1</sub> population: recovery of RP genome

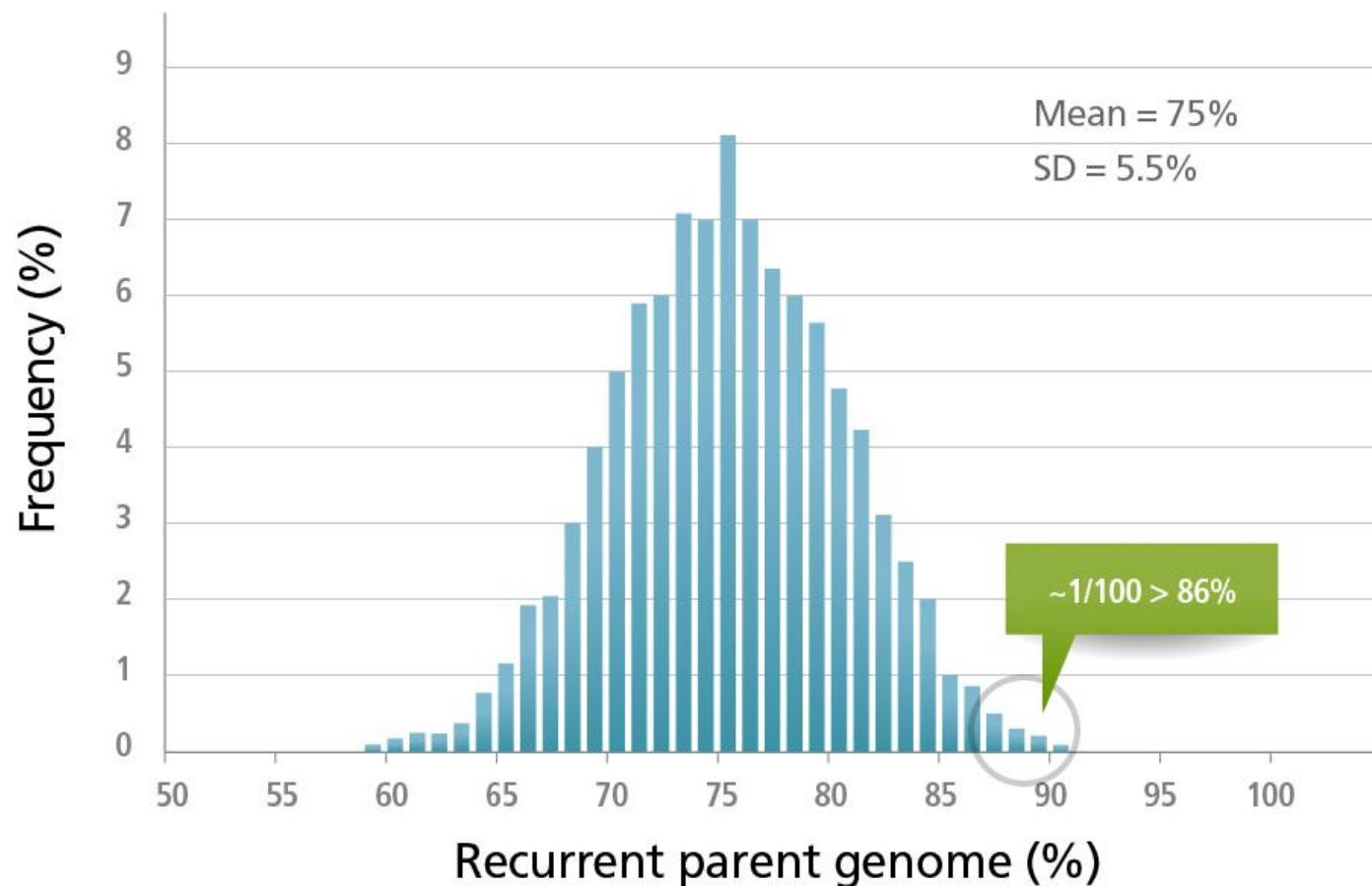


Figure Credit: Modified from Welz and Geiger, 2000



# American chestnut

- Early in the 20th century, the Appalachian Mountains were full of giant chestnut trees
- Chestnuts were abundant, accounting for 25% of all Appalachian hardwoods
- Tree diameters of 8 to 10 feet were often reported. One in Francis Cove, North Carolina, was 17 feet in diameter
- Chestnut grew tall (up to 120 feet) and straight -- often clear of branches up to 50 feet, making them ideal lumber



Image Credit: John Carlson, The Pennsylvania State University and The American Chestnut Foundation.

# American chestnut



Image Credits: John Carlson, The Pennsylvania State University and The American Chestnut Foundation

# American chestnut

- Chestnut was an important commodity for the early European settlers of the Appalachians
- Uses included
  - *Nuts for food, mast, and cash*
  - *Tanning hides*
  - *Building materials*

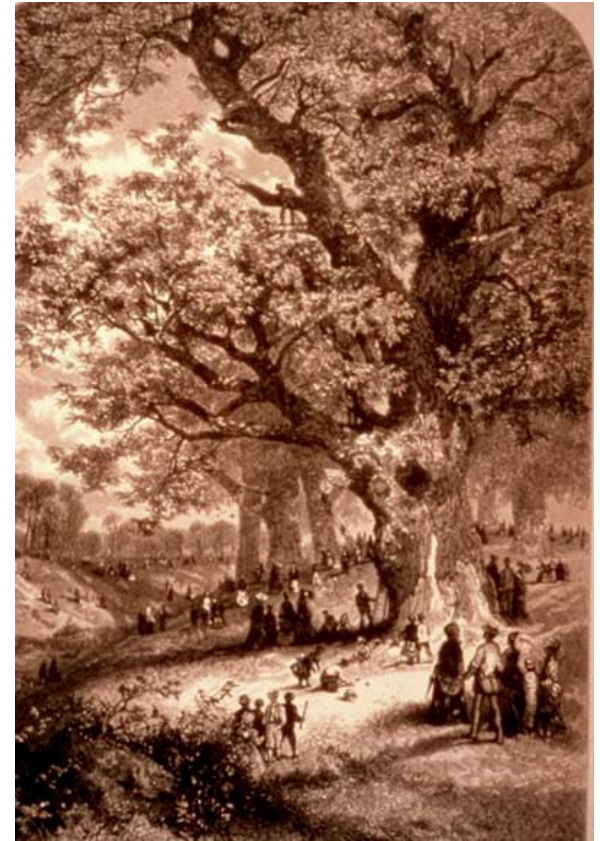
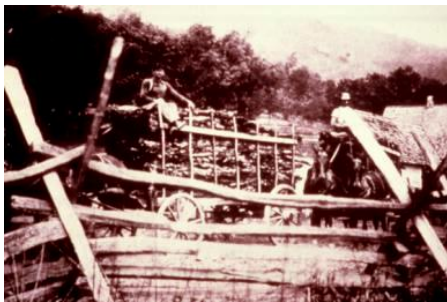


Image Credits: John Carlson, The Pennsylvania State University and The American Chestnut Foundation



# The chestnut blight

- By the 1950s, chestnut was virtually eliminated as a dominant forest tree



Image Credits: John Carlson, The Pennsylvania State University and The American Chestnut Foundation

# QTL studies reveal three sources of major resistance

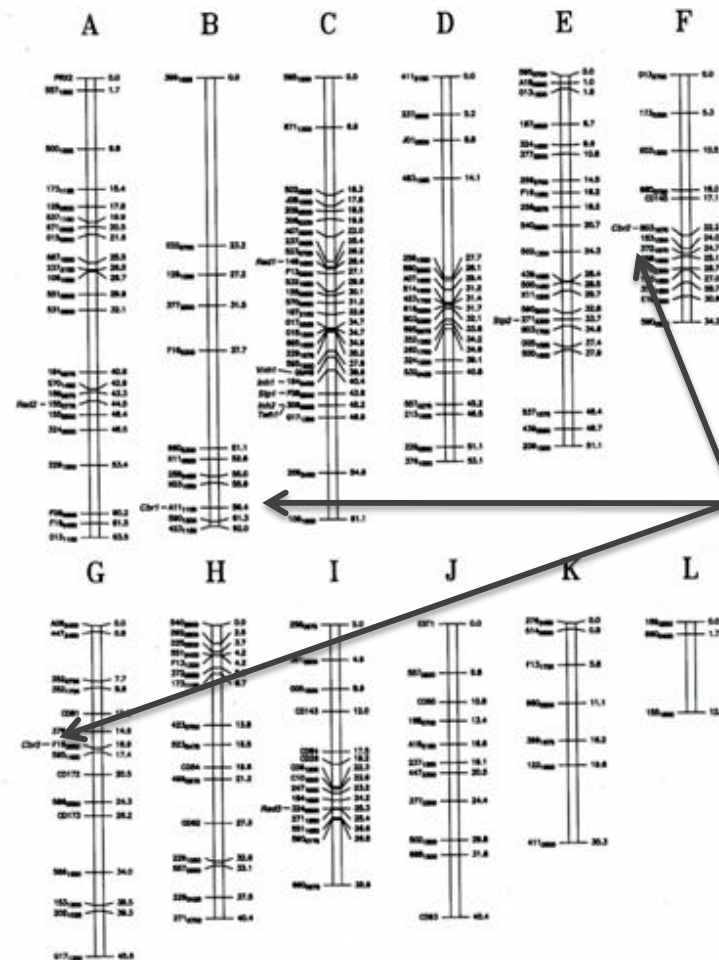


Figure Credit: Modified from Kubisiak et al., 1997

# Marker applications in chestnut restoration

- Quality control in breed orchards
- Genetic mapping
- QTL mapping
- Marker informed introgression
- Locating resistance genes



Image Credits: Brad Smith and Fred Heberd, The American Chestnut Foundation

# Propagation population characterization

- Varietal protection (quality control)
- Parentage analysis / orchard efficiency
  - *Pollen contamination*
  - *Parental contribution to the gene pool*
  - *Pollen competition*
  - *Supplemental mass pollination success*
  - *Mating systems*

# Pollen contamination

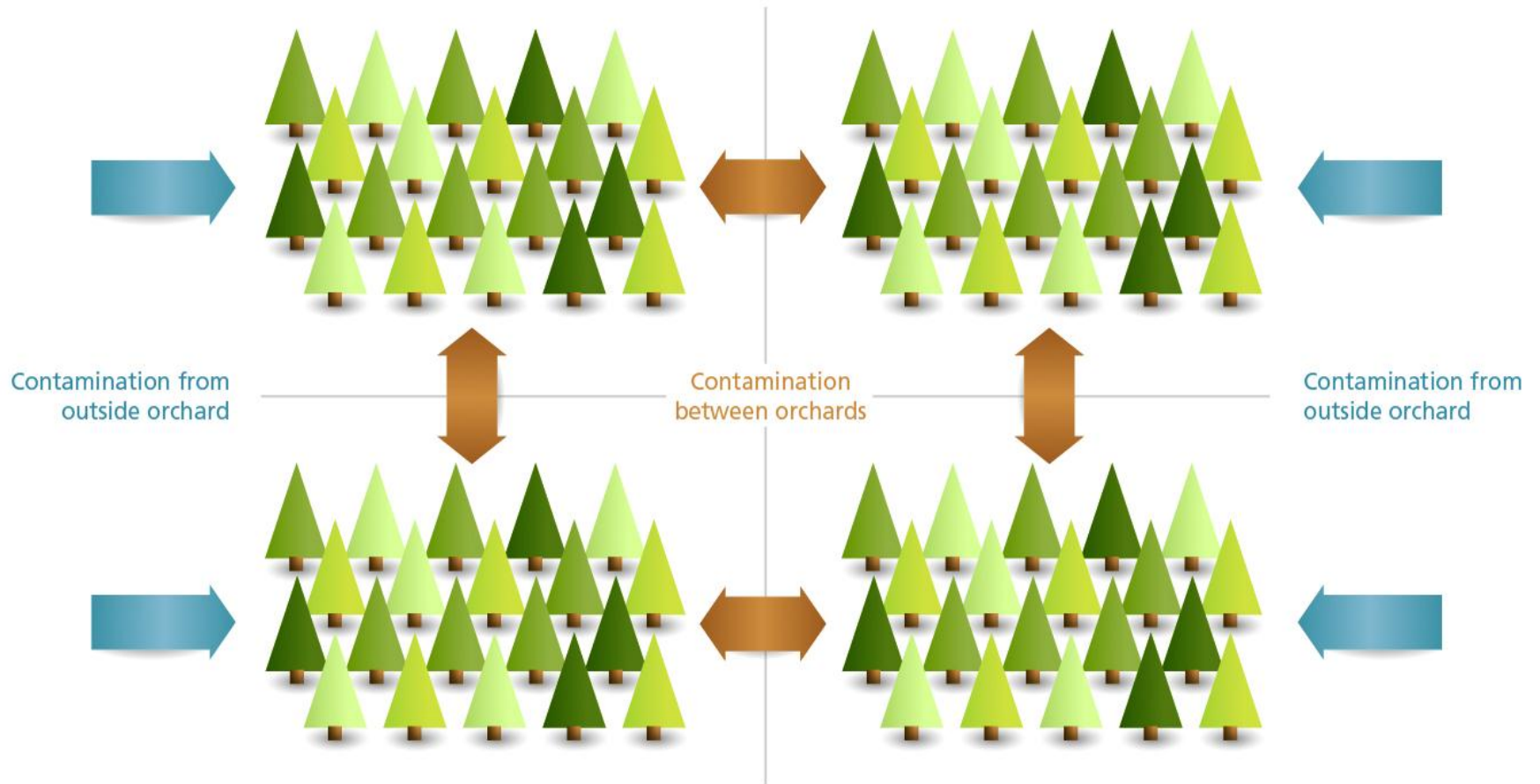


Figure Credit: Nicholas Wheeler, Oregon State University



# Pollen contamination in DF orchards

Year	Type of seed collection	# of seeds analyzed	Observed seed contamination (%)	Pollen contamination (%) $\pm$ SE
1999	Bulk	192 (190 <sup>b</sup> )	1.0	31.0 $\pm$ 3.5
2000	Bulk	192 (102 <sup>b</sup> )	46.9	36.8 $\pm$ 5.2
2000	Individual-ramet	240	0	32.0 $\pm$ 3.2
2003	Individual-ramet	336	0	41.3 $\pm$ 2.8
<b>Mean</b>				<b>35.3 <math>\pm</math> 2.4</b>

Slavov et al., 2005; Figure Credit: Glenn Howe, Oregon State University

# PC is higher in parents that flower early

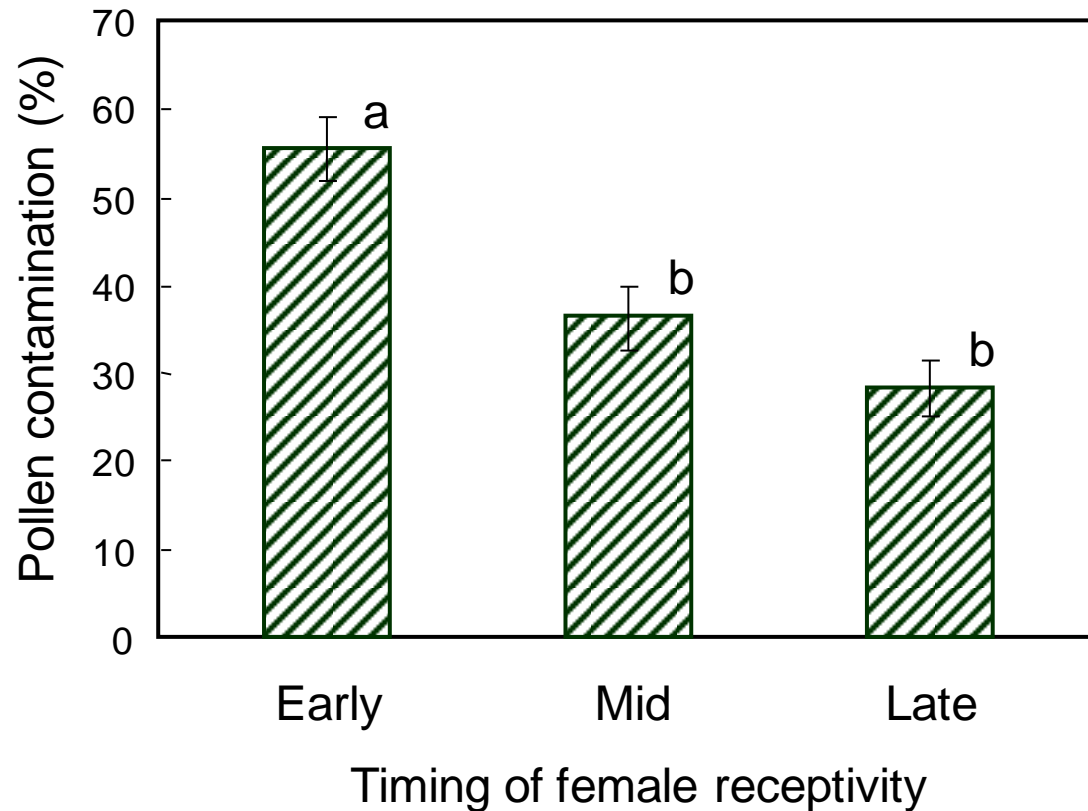


Figure Credit: Glenn Howe, Oregon State University

# Differential paternal success

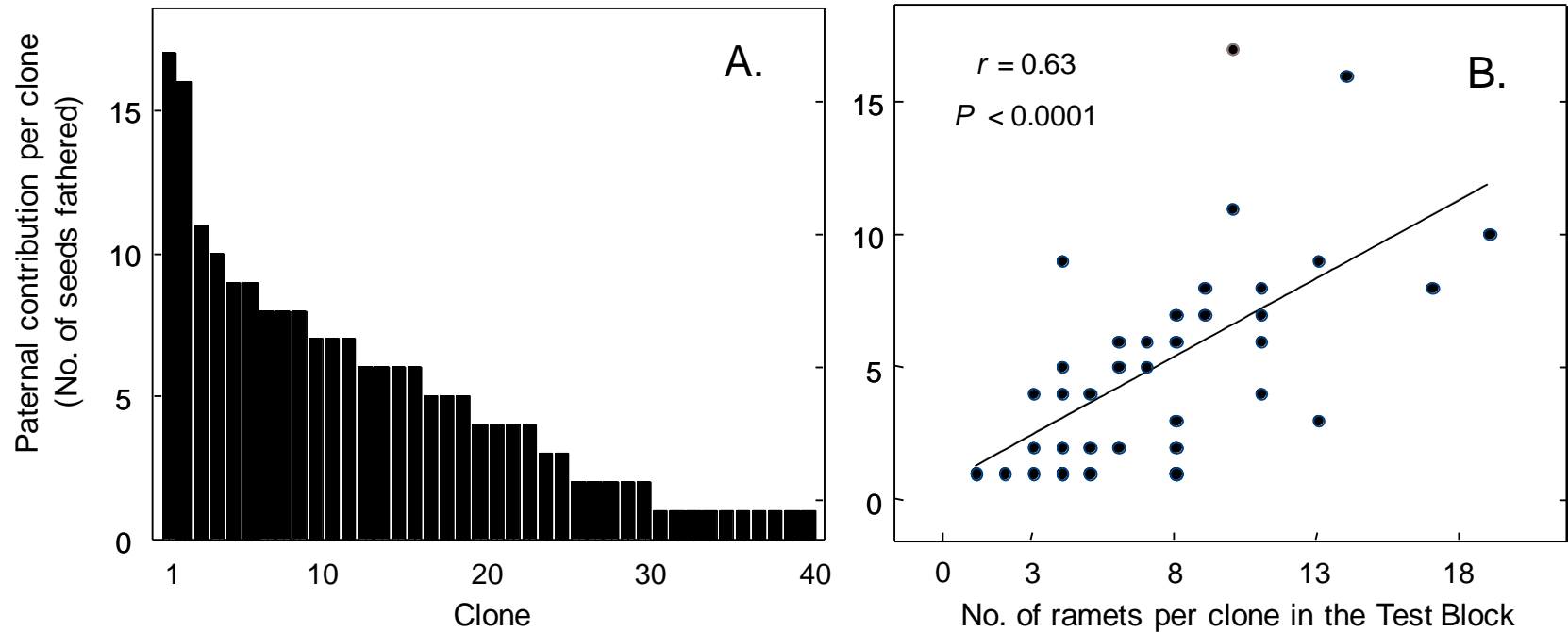


Figure Credit: Glenn Howe, Oregon State University

# Parentage reconstruction

**Table 5.** Parent tree identification using SSR genetic markers. The SSR genotypes of two putative open-pollinated (OP) field selections were compared to the genotypes of known progeny growing in genetic test plantations (n = 8-10). If the putative parents in the field are correct, then all of their progeny must have at least one of the two parental alleles. Cases in which a progeny allele matches one of the alleles in the putative parent are shown in white. Allele numbers (e.g., 209 or 216) represent relative lengths of alternative SSR alleles.

Putative OP parent (i.e., tree in the field)		Progeny number (in progeny test plantation)											Inferred genotype of real OP parent	Conclusion: putative parent is:
ID	SSR genotype (marker name)	Progeny allele	1	2	3	4	5	6	7	8	9	10		
1	209, 216 (OSU_3F1)	Shorter = Longer =	205 216	209 209	202 209	216 216	209 219	216 227	216 223	209 216	209 211	216 218	209, 216	Correct
1	184, 191 (OSU_3B9)	Shorter = Longer =	172 191	188 191	184 184	184 184	176 184	172 191	191 191	191 191	186 191	184 191	184, 191	Correct
1	254, 256 (OSU_4A7)	Shorter = Longer =	254 254	254 284	228 256	256 284	226 256	226 254	254 254	254 254	226 254	254 254	254, 256	Correct
2	204, 222 (OSU_3F1)	Shorter = Longer =	210 224	210 214	192 210	210 214	210 226	214 214	190 210	214 220			210, 214	Incorrect
2	130, 190 (OSU_3B9)	Shorter = Longer =	130 208	188 194	130 210	130 196	188 200	130 130	130 188	130 190			130, 188	Incorrect
2	226, 226 (OSU_4A7)	Shorter = Longer =	226 226	226 228	226 244	226 248	226 284	268 284	228 284	282 284			226, 284	Incorrect

Figure Credit: Glenn Howe, Oregon State University

# Number of loci needed for paternity in DF

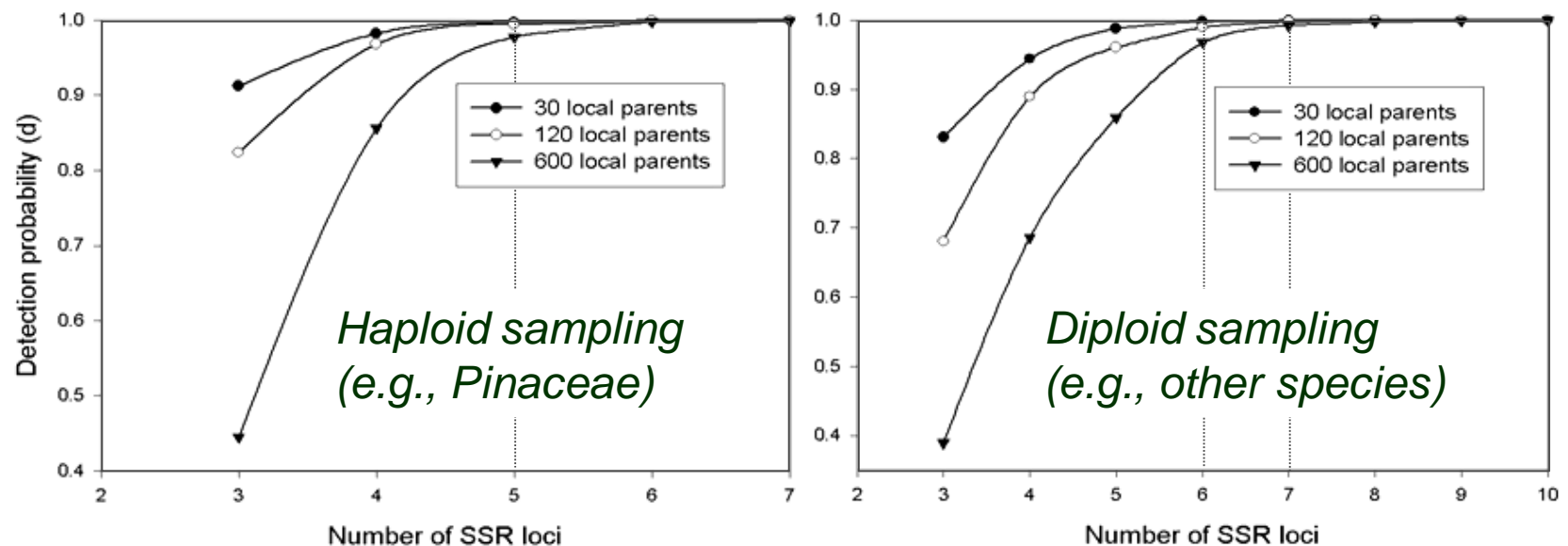


Figure Credit: Glenn Howe, Oregon State University

# Enhanced breeding designs: Informed choices for full-sib matings

- Complementary breeding
  - *Identify individuals that differ (genotypes) from one another at a key locus (i.e. R genes for disease resistance)*
  - *Mate to distribute R genes into other backgrounds*
- Pyramiding genes
  - *An extension of complementary breeding where you attempt to accumulate desirable alleles at two or more loci into one progeny cohort (see example later)*
- Diversity index breeding

# Enhanced breeding designs: Using markers for paternity analysis

## Polymix breeding with paternity analysis (PMX/WPA)\*

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- Replace multiple breeding designs (PMX for BV estimation + Full-sib design for advanced generation selection) with just a single PMX test
- Fingerprint the top individuals in the PMX test to determine paternity (maintains pedigree control and manages inbreeding)
- Saves time and cost of full-sib breeding while actually increasing gain potential

\* Lambeth, et al., 2001

# Tree improvement flow diagram

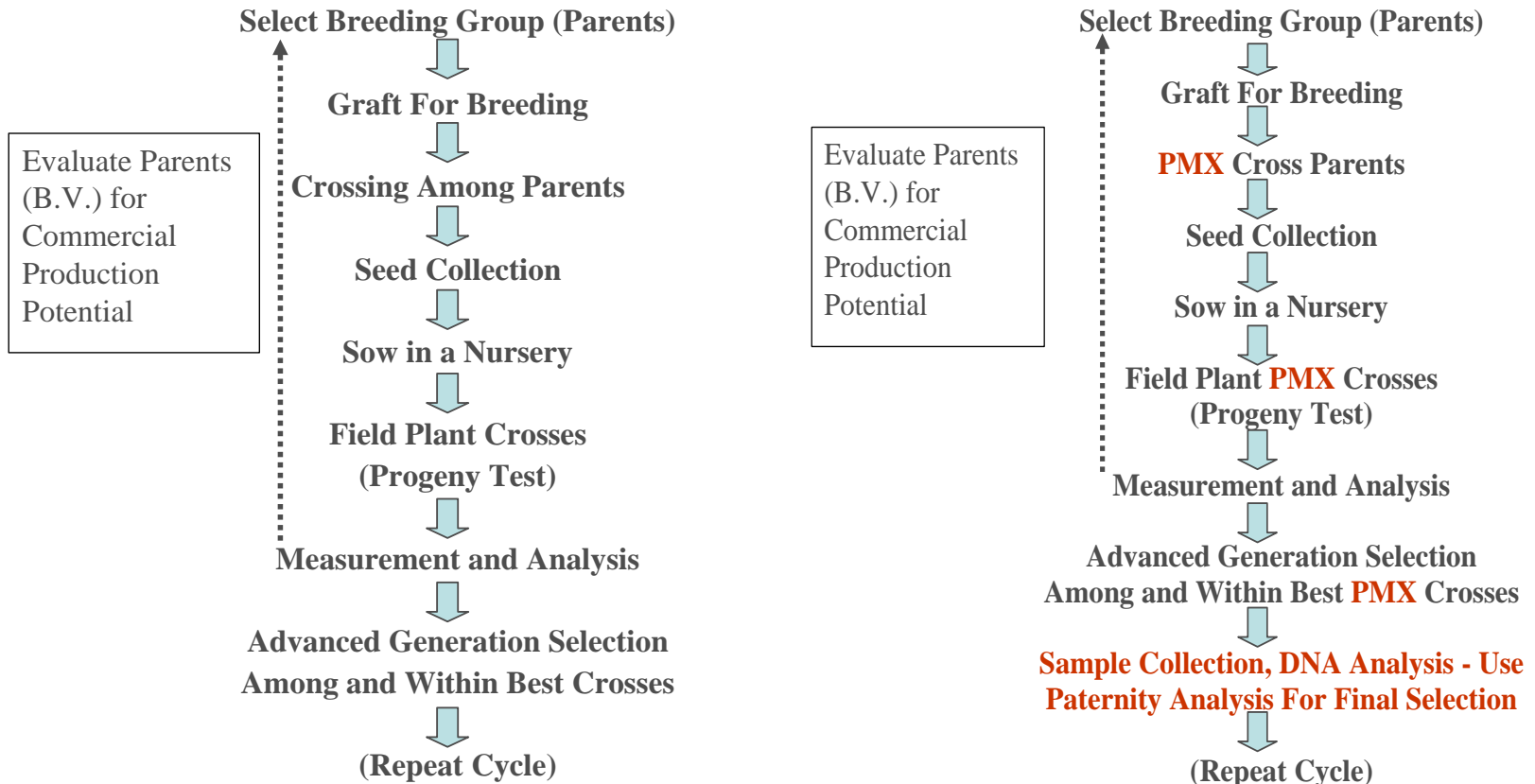


Figure Credit: Clem Lambeth



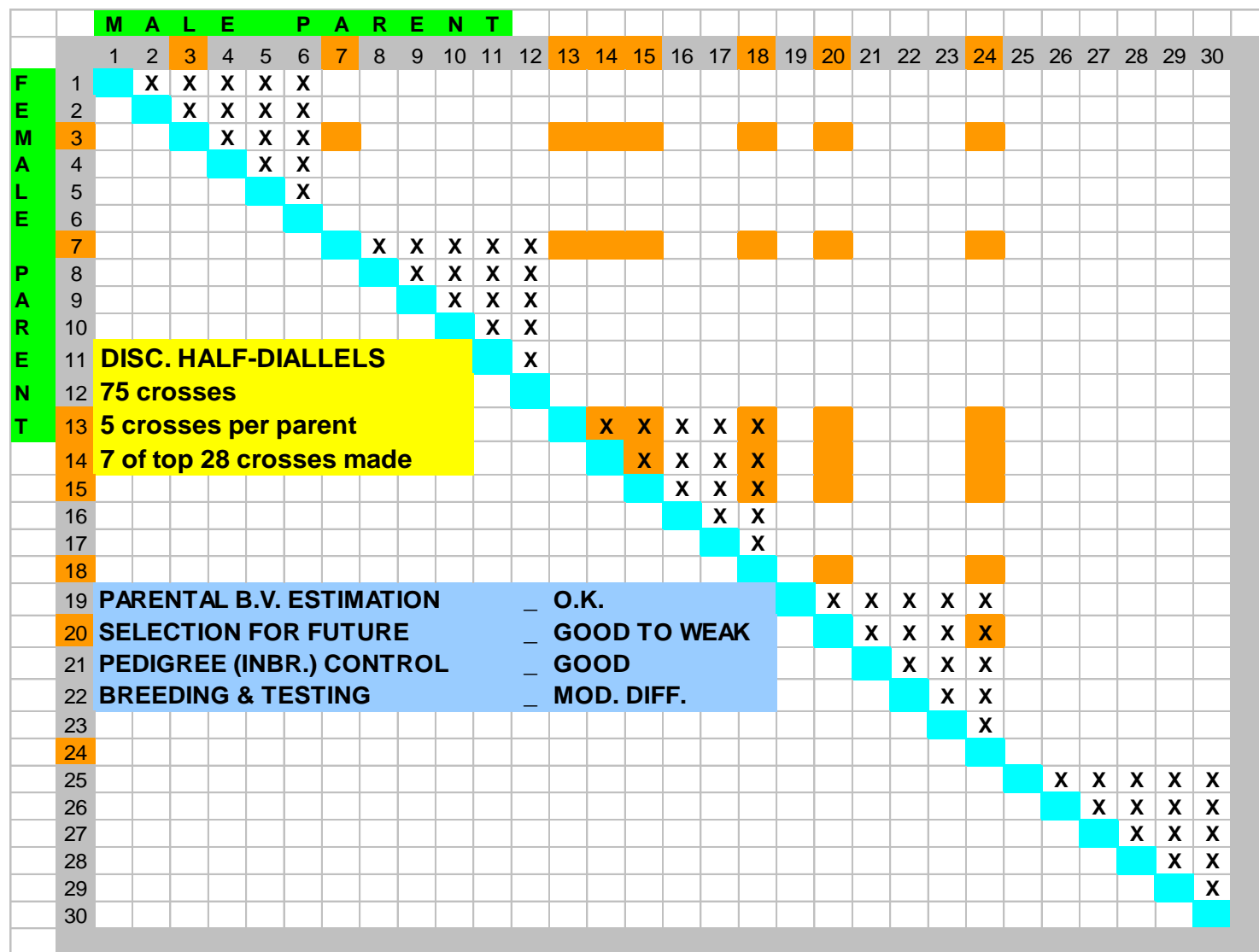


Figure Credit: Clem Lambeth

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Retrospective selection strategy of elite parents based on paternity testing of progeny individuals displaying superior performance using microsatellite markers.

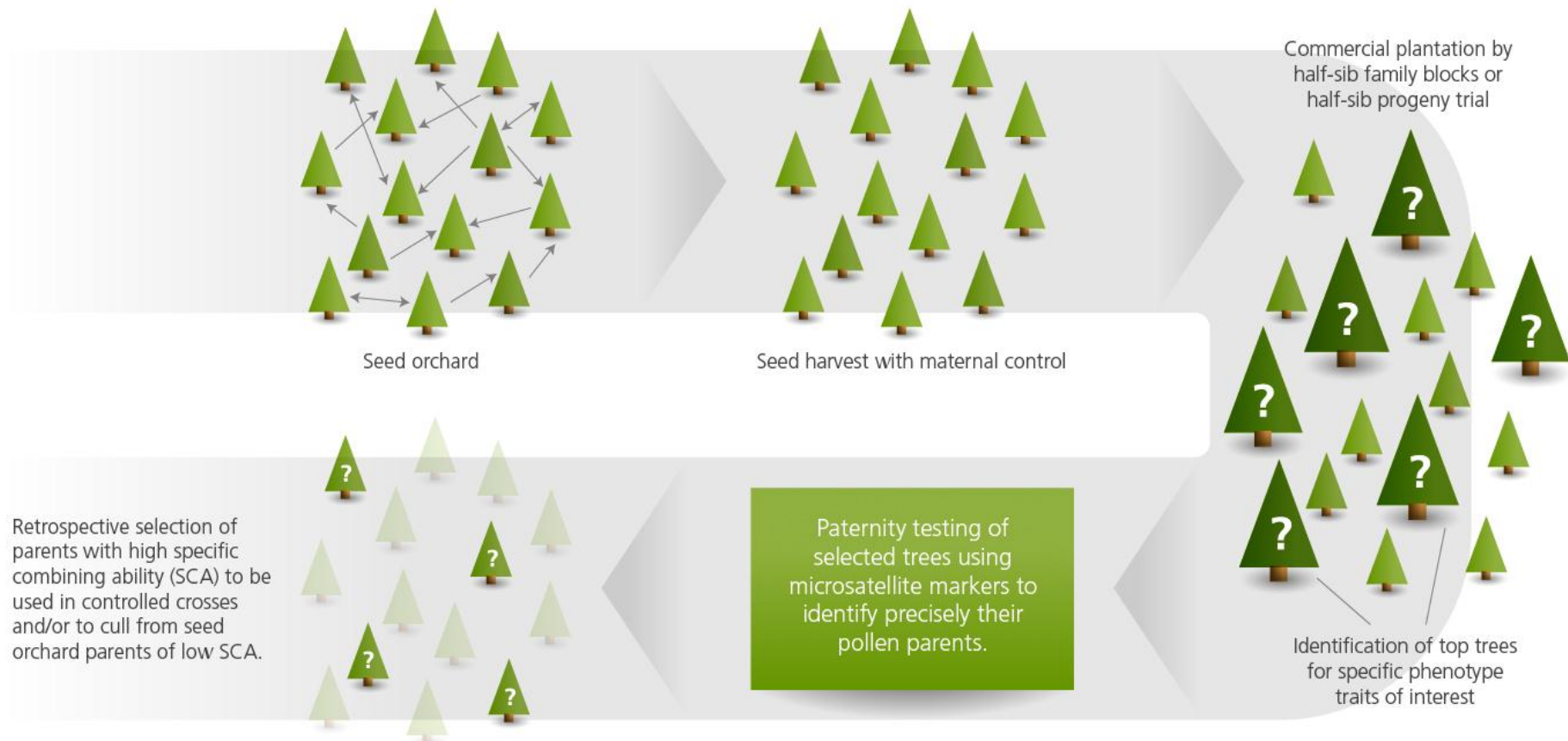


Figure Credit: Modified from Grattapaglia, 2007

# Implementation strategies for marker informed breeding in tree improvement

- “All forms of MAS can be applied separately or in conjunction with classical methods of selection (mass, family, within-family, combined and index selection) and can be utilized to make selections for selected, breeding and/or production populations”
  - *In White, Adams and Neale, Forest Genetics, Chapter 19, Page 554*

# Approaches to MAS: (Classified by mapping precision)

Classification of three different types of marker-trait associations relevant to *Eucalyptus* MAS  
(see text for details)

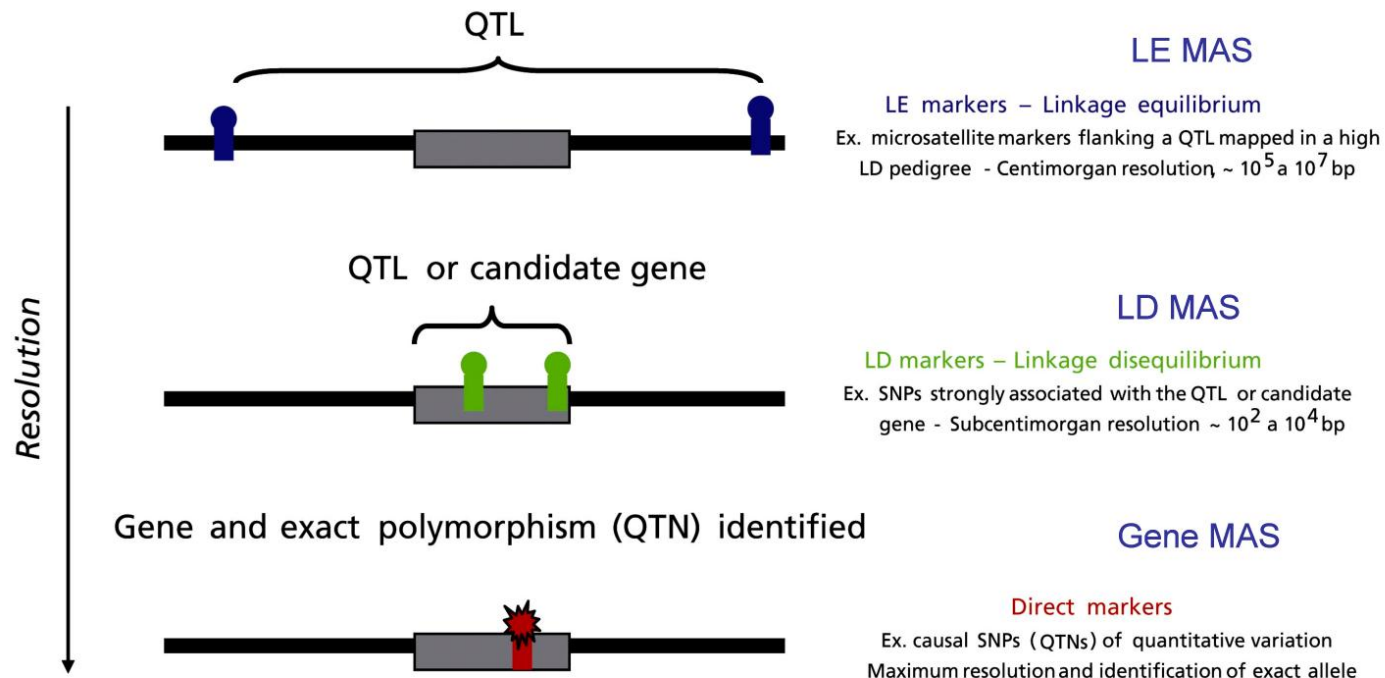
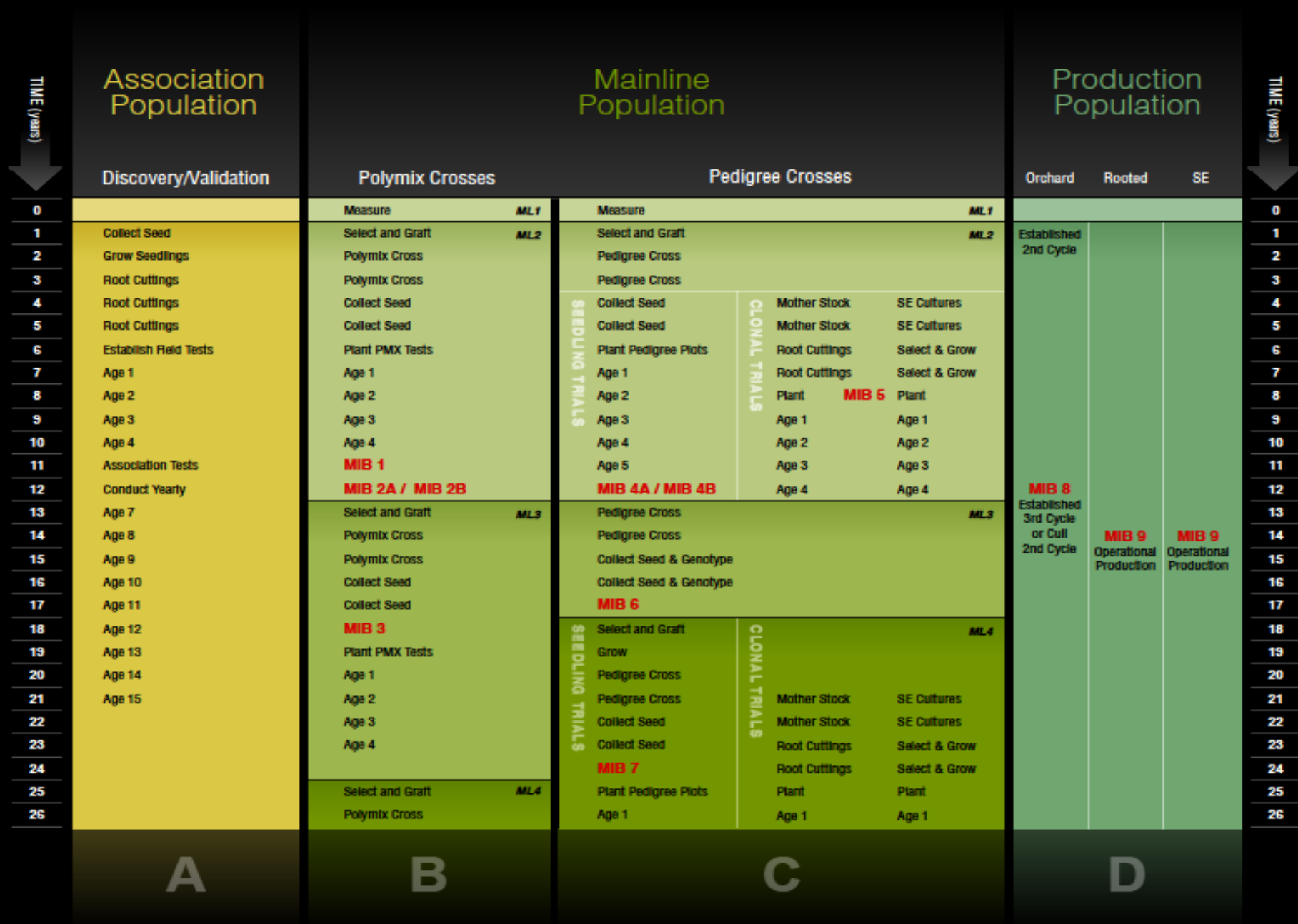


Figure Credit: Modified from Grattapaglia, 2007



# References cited

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# External link

- Forest health initiative. [Online]. Available at:  
<http://foresthealthinitiative.org> (verified 2 June 2011).



# Thank You.

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