Using Bioinformatics to Identify Promoters in Genome Sequences

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Purpose of this tutorial

☐ Provide step-by-step instructions to automatically identify DNA sequences in a genome sequence using a BioPerl script

Rationale

- ☐ With the continual release of plant genome sequences, the accumulation of genomics information has been exponentially increasing
- □ Automated analysis and mining of genome databases is becoming essential and routine due to the volume of data

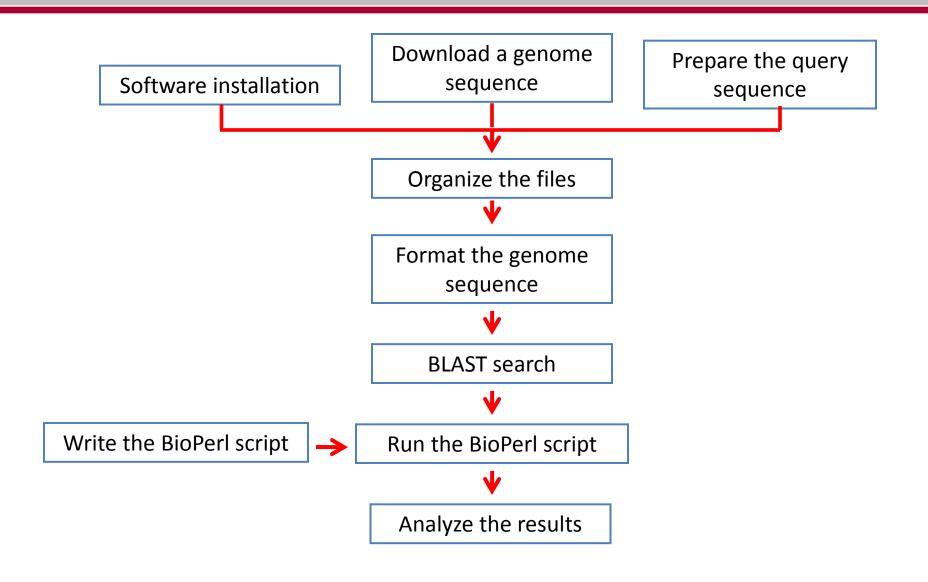
Biological rationale for targeting promoters

☐ Promoters are DNA sequences located in front of gene coding sequences ☐ Due to their influence on gene expression, promoters are of interest as tools to regulate Genetically Modified Organism (GMO) products and as markers for natural variation ☐ Promoters with different functionality in terms of inducibility, tissue specificity, strength etc. are needed ☐ In this tutorial, we will focus on the identification of ubiquitin promoters of tomato which drive strong gene expression in important crops such as maize (Christensen and Quail 1996), rice (Sivamani and Qu 2006), potato (Garbarino et al. 1995), tomato (Rollfinke et al. 1998) and soybean (Hernandez-Garcia et al. 2009)

Specific objective:

To identify *ubiquitin* genes and their promoter sequences in a draft genome of tomato using a BioPerl script in a Windows operating system

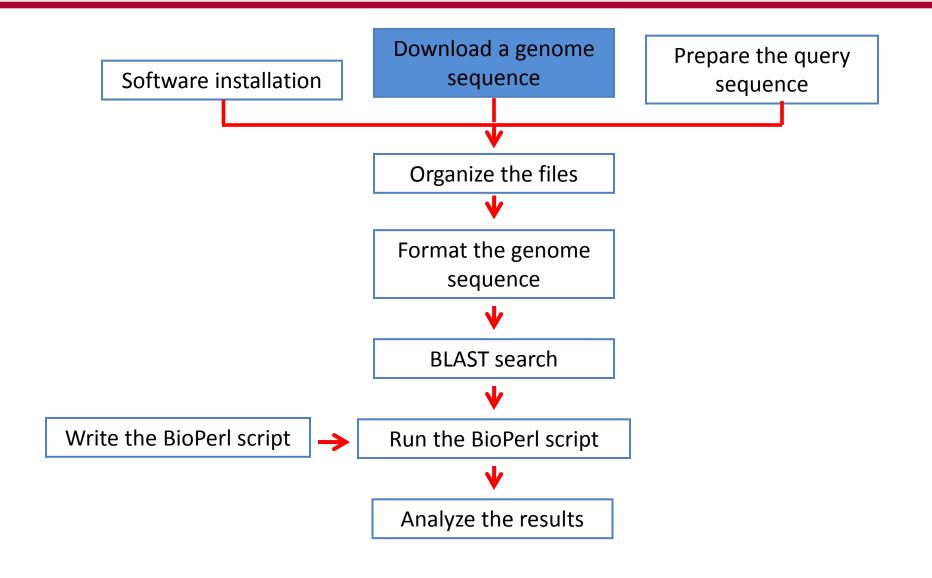
Methodology



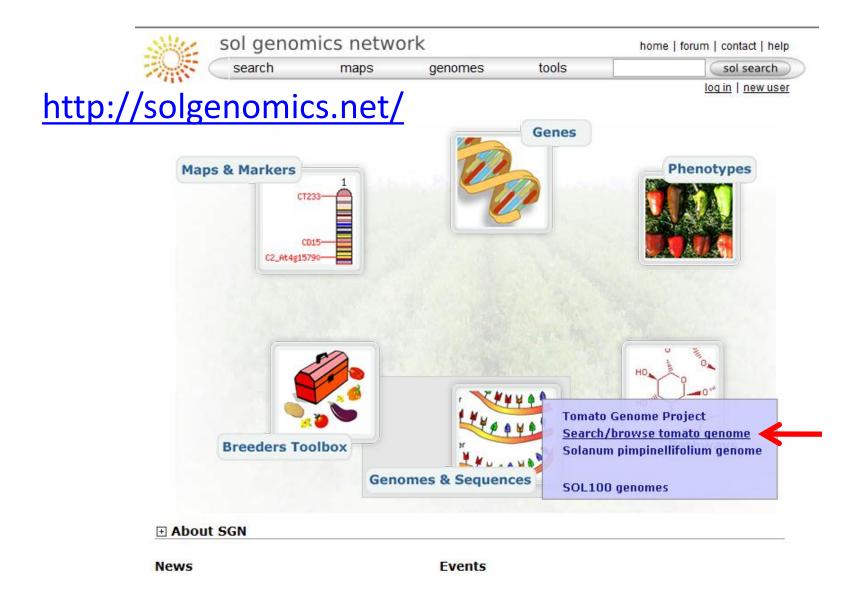
1. Software installation

☐ Cygwin, UNIX emulator for Windows OS http://www.cygwin.com/ ☐ BioPerl http://www.bioperl.org/wiki/Main Page ☐ StandAlone BLAST http://www.ncbi.nlm.nih.gov/staff/tao/URLAPI/pc_setup. html □7-ZIP compressor software http://www.7-zip.org/

Methodology



2. Download a draft genome sequence of tomato



2. Download a draft genome sequence of tomato

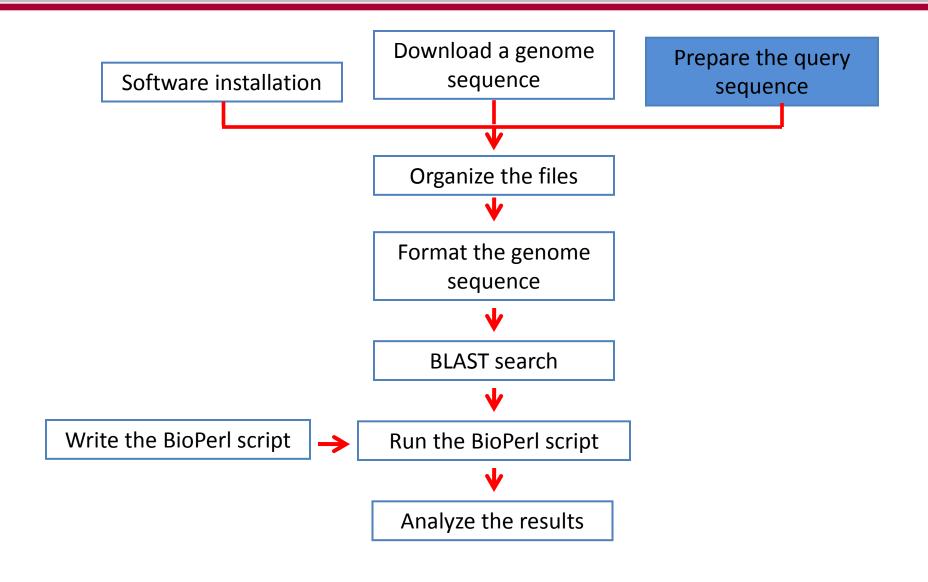
http://solgenomics.net/



Tomato Genome Data

To	Tomato genome sequence builds								
	Release	Date	Description	Annotation	Download				
	1.00	Dec 2009	initial build, based on the Newbler assembler and containing only 454 sequencing data	ITAG1	scaffolds proteins cds				
	1.03	Jan 2010	like 1.00, but with additional 454 runs and improved contamination screen	Not annotated	scaffolds				
	cabog1.00	Mar 2010	All 454 data, bac end and fosmid end data, assembled using the CABOG assembler	Not annotated	scaffolds				
	1.50	Apr 2010	Includes all 454 data, bac ends, fosmid ends, polishing with Solexa and SOLiD data	Not annotated	scaffolds				
	2.00	Jun 2010	Release withdrawn.	Not annotated	-				
	2.10	Jun 2010	Additional scaffold merging using clone end sequences. Scaffolds placed and oriented using multiple physical maps, first release to include chromosome pseudomolecule sequences.	Not annotated	scaffolds, chromosomes				

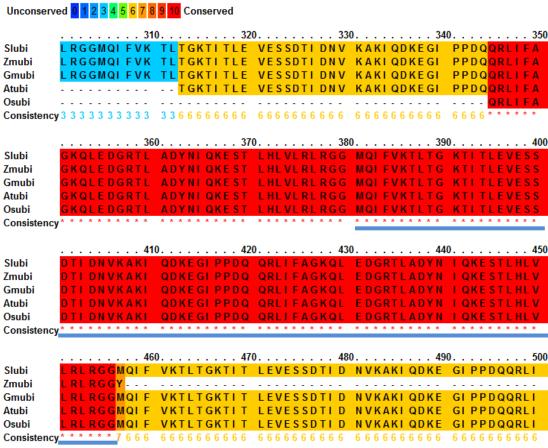
Methodology



3. Preparing the query sequence

The query sequence used in this tutorial corresponds to an ubiquitin domain present in ubiquitin proteins. Note that ubiquitin proteins are highly conserved in plants and contain single or multiple ubiquitin domains. This is what allows us to look for gene sequences in tomato using information from other plant species. The figure below depicts an alignment of ubiquitin proteins from tomato (Slubi, GenBank: CAA51679.1), maize (Zmubi, GenBank: AAC49014.1), rice (Osubi, GenBank: BAA02241.1), *Arabidopsis* (Atubi, GenBank: ABH08755.1) and soybean (Gmubi, GenBank: BAA05085.1).

Sequences were aligned using the PRALINE multiple sequence alignment tool (http://www.ibi.vu.nl/programs/pralinewww/). A complete ubiquitin domain (75 amino acid) is underlined.



3. Preparing the query sequence

For simplicity, the nucleotide sequence from soybean corresponding to the 76-amino acid domain from ubiquitin was used as the query sequence.

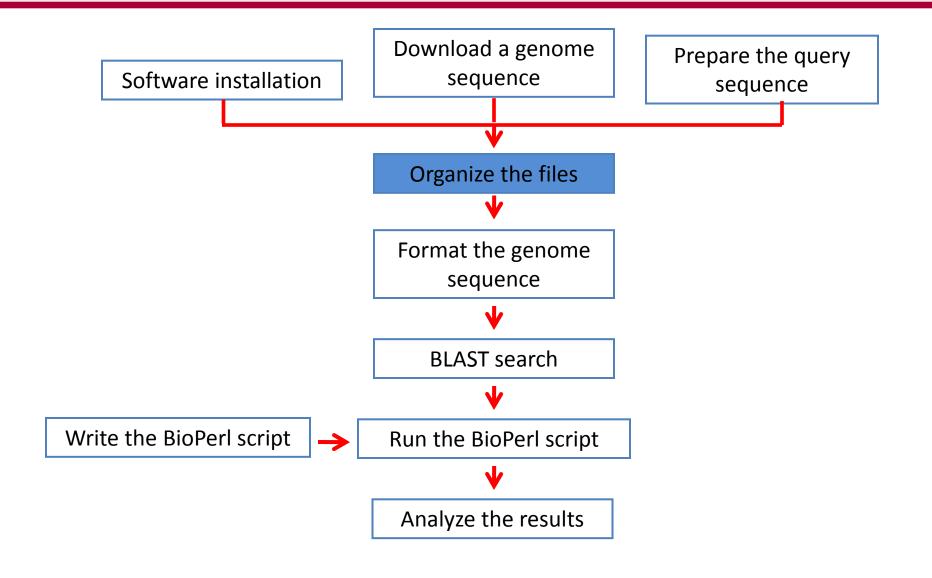
The nucleotide sequence, in the fasta format, was placed in a ".txt" file using a text editor.



>Gmubidomain

ATGCAGATCTTCGTCAAGACCCTCACCGGCAAGACCATCACCCTTGAGGTGGAAAGCTCTGACACCATCGA CAACGTCAAGGCCAAGATCCAGGACAAGGAAGGAATCCCCCCGGACCAGCAACGTCTCATTTTCGCCGGAA AGCAACTTGAGGACGGCCGTACCCTTGCTGACTACAACATTCAGAAGGAGAGTACTCTTCACCTCGTCCTC

Methodology



4. Organizing the files

Decompress the genome sequence file with any compressor software (e.g. 7-ZIP compressor)

Place your decompressed genome sequence file (*SL2.10sc*) and the query sequence file (*Gmubidomain*) in a new created folder (e.g C:\cygwin\home\Carlos*Solanum_Lycopersicum*)

5. Format the genome sequence before using BLAST

Open UNIX (In our case, we are using Cygwin emulator for PC) Go to Carlos@Carlos-PC ~/Solanum_Lycopersicum Type formatdb.exe -i SL2.10sc.fasta -p F -i indicates the input file to format into a searchable database

- -p asks if the input data is protein sequence

F indicates "false", which specifies a nucleotide database

```
arlos@Carlos-PC
 cd Solanum_Lycopersicum
Carlos@Carlos-PC ~/Solanum_Lycopersicum
 formatdb.exe -i SL2.10sc.fasta -p
```

5. Format the genome sequence before using BLAST

Output files

Name	Date modified	Туре	Size	Ta
SL2.10sc	6/25/2010 8:53 AM	FASTA File	772,567 KB	
formatdb	8/15/2010 9:20 PM	Text Document	1 KB	
SL2.10sc.fasta.nhr	8/15/2010 9:20 PM	NHR File	259 KB	
SL2.10sc.fasta.nin	8/15/2010 9:20 PM	NIN File	41 KB	
SL2.10sc.fasta.nsq	8/15/2010 9:20 PM	NSQ File	197,554 KB	
Gmubidomain	8/5/2010 3:38 PM	File	1 KB	
SL_Gmubi	8/5/2010 3:40 PM	OUT File	40 KB	
🎉 find_blast_matches	8/5/2010 4:26 PM	PL File	8 KB	

A correct format will create three files (NHR, NIN and NSQ files). In addition, you will see a report for completion in a TXT format.

6. BLAST the query sequence against the genome sequence

Open Cygwin
Go to Carlos@Carlos-PC ~/Solanum_Lycopersicum

Type blastall.exe –p blastn –d SL2.10sc.fasta –i gmubidomain –o SL_Gmubi.out

- **-p** is the name of the program to use (**blastn**)
- -d specifies the file for the genome sequence (SL2.10sc.fasta)
- -i indicates the query sequence file (gmubidomain)
- -o indicates the name of the output file that will be created (Gmubi.out)

7. Checking the output file

Query: 61

In this file, we expect to see a list of sequences with significant alignments and their e-values, and the actual alignments between the query sequence and the subject genome

```
SL_Gmubi - Notepad
File Edit Format View Help
BLASTN 2.2.20 [Feb-08-2009]
Reference: Altschul, Stephen F., Thomas L. Madden, Alejandro A. Schaffer,
Jinghui Zhang, Zheng Zhang, Webb Miller, and David J. Lipman (1997),
"Gapped BLAST and PSI-BLAST: a new generation of protein database search
programs", Nucleic Acids Res. 25:3389-3402.
Query= Gmubidomain
         (228 letters)
Database: SL2.10sc.fasta
          3433 sequences; 781,288,557 total letters
                                                               Score
Sequences producing significant alignments:
                                                               (bits) Value
SL2.10sc05010
                                                                    180
                                                                          6e-044
5L2.10sc04626
                                                                    180
                                                                          6e-044
                                                                    172
SL2.10sc05925
                                                                          1e-041
SL2.10sc05380
                                                                    125
                                                                          3e-027
                                                                    103
SL2.10sc03748
                                                                          1e-020
                                                                     72
SL2.10sc04323
                                                                          4e-011
SL2.10sc03806
                                                                          9e-009
SL2.10sc06101
                                                                          1e-007
                                                                          4e-005
SL2.10sc05732
SL2.10sc03923
                                                                          1e-004
SL2.10sc04813
                                                                          0.002
                                                                     40
5L2.10sc06214
                                                                          0.13
SL2.10sc05632
                                                                     36
                                                                          2.1
SL2.10sc04126
                                                                          2.1
                                                                     36
SL2.10sc03796
                                                                          2.1
SL2.10sc04777
                                                                          8.3
SL2.10sc03714
                                                                     34
                                                                          8.3
SL2.10sc03685
>SL2.10sc05010
         Length = 20453289
Score = 180 bits (91), Expect = 6e-044
Identities = 193/227 (85%)
Strand = Plus / Minus
Query: 1
               atgcagatcttcgtcaagaccctcaccggcaagaccatcacccttgaggtggaaagctct 60
               Sbjct: 13588607 atgcagatcttcgtgaaaaccctaacggggaagacgatcaccctagaggttgagtcttcc 13588548
```

8. BioPerl script

The BioPerl script used in this tutorial (provided as a .txt file, do not forget to change the file extension to .pl):

- -Parses the output blast file against the genome sequence file to identify the sequences with the highest similarities with the query sequence
- -Extracts the promoter sequences for those genes
- -Allows the adjustment of e-values and the length and the number of extracted promoters

```
find blast matches - WordPad
File Edit View Insert Format Help
#!/usr/bin/env perl
 # Made by Gabriel Abud
 # This script takes a blast output file and a contig or genome file as
 arguments (in that order).
 # It then parses through the blast file to check for good matches
 # Those matches are then used back into the genome file, and the
 sequence in the genome
 # is displayed (to STDOUT)
 # To redirect the output to a file, use the Unix '>' operator
 # Note: If you want to see the output in a Unix shell, please be sure
 to maximize the terminal
 # before hand.
 # Changes:
 # Optional flags for maximum e-value, amount of hits shown, and
 promoter region were added
 # Use --help for more information
 use warnings;
 # Modules
 use lib '/cygdrive/c/Perl/site/lib';
 use lib '/cvgdrive/c/Perl/site/lib/Bio';
 use Bio::SearchIO;
 use Bio::Seg;
 # use File::Basename;
 # Variables
 my @scaffolds;
 my $inputFile;
 mv $scaffoldFind:
```

The command --help displays a brief description of the main parameters that can be adjusted:

- **-e** to modify the e-value (the default is 0.01)
- -p to specify the promoter length
- **-n** to specify the number of promoters (the default is one promoter at a time)

The desired values are entered after the specification of each parameter

```
# Help screen
52

    if( defined($ARGV[0]) && "$ARGV[0]" =~ /^-?-?help$/i ) {
          print "\n$baseProg:\n\n\n";
54
          print "Syntax:\n\t$baseProg blast_output_file contig_or_genome_file [OPTIONS]\n\n";
55
56
          print "Options:\n";
57
          print "\t-e, maximum e-value for matches (0.01 by default)\n\n";
          print "\t-p, base pairs of promoter region to be included (should only be used in DNA sequences)\n\n";
58
          print "\t-n, number of top hits to display, starting with the highest hit (1 by default)\n\n";
59
          exit
```

Commands to enter to execute the script

```
# Checks for correct amount of arguments The blast file

if ( @ARGV < 2 ) {

print STDERR "USAGE: $baseProg blast output file contig or genome file [OPTIONS]\n";

print STDERR "Type $baseProg --help' for more information\n";

exit;

The parameters: -e -p -n

The script's name
```

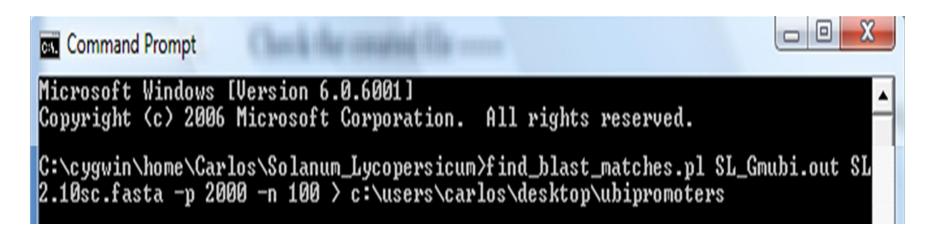
Parameters specified by default

This script is able to extract promoters in the + and – strands. This is done by getting the reverse complement of - strands

9. Run the BioPerl script

Now we need to extract the upstream sequences (from our genome sequence) for genes with high similarities identified during the previous Blast procedure.

- -Go to Command Prompt
- -Locate the main folder containing all the files (Solanum_Lycopersicum folder)
- C:\cygwin\home\Carlos\Solanum_Lycopersicum>
- -Type: find_blast_matches.pl SL_Gmubi.out SL2.10sc.fasta -p 2000 -n 100 > c:\users\Carlos|Desktop\ubipromoters
- -p is promoter length (2000 bp)
- -n 100 pulls out up to 100 promoters if present (the default is 1 promoter)
- The default e-value is 0.01
- ">" indicates the creation of an output file (txt) at any specified location



10. Analyzing the output file

☐ The output file contains the promoter sequences identified in our target genome □31 ubiquitin promoters were found in the draft genome sequence of tomato ☐ Each promoter is extracted from an *ubiquitin* gene with at least one ubiquitin domain, therefore, 31 *ubiquitin* genes were identified in this draft genome of tomato

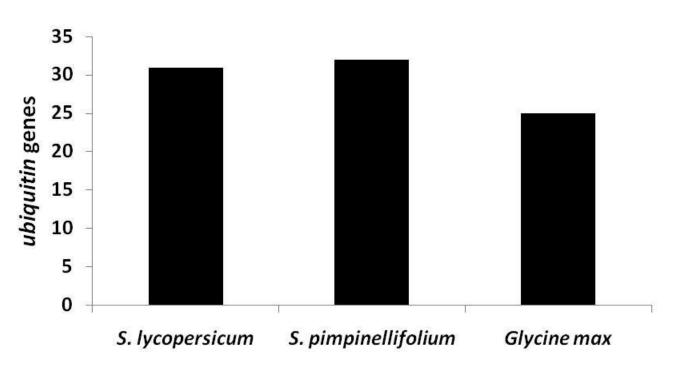
ubipromoters - Notepad

File Edit Format View Help

>SL2.10sc05925 (SL2.10sc.fasta) at 4246810 - 4247037 Promoter TCAAGAGTTTTTTCATACACACACACTTTAGGCTCAAAATCAAGATCTTTGATTAAGAAAATAATAGTTTC GTCTACAACCCACTATCATTTAGTGCTCAAATATTGCTTCTTTTTCTTCTTATCAGTTGAAGTTAGATTT TTCAGTTTAAAATTTTAGAACATCGTGCAATTGAGTCCATTTTATTCAATTCAAAGTTTAAGATT CTATAGCATCGTAAAGTTTAATTTTTTAAGTTTATACCTTAGAGATATTGTATCCTAAAATCGAGTTGTG TGATTCGAACTTACTATTGGTAGAATAATTATTTTGATTAACCATCAGATGAAGTTGCCCCTTTCATAAC ACATATCACGATTAAGTACGAAAAATATTTCCTAACATAATTTAAATTAGTCGAATTCTAACGAATATCA CACCTAAATAGATCTCACGTGATAGAAATTTGACTTAGTCAGACTCCAATACTCGCTCAAAAAT. TATATAGGAAGTTGCCATCCAAACGGCACCTAAATTTTCTCCACCGATCAAAGAAAAGGAGAGCG ACTATTATTGGTTACTTCATTGGCAATTTCTTCACAAATTGGCCATTAACAACTATAAAAGGAGG CTTTTCCTTCTTTTAGTTTTTTTGTTTATTTTTTCTATATTTTGTGCTATTTGTTTTGTTGTG TTTAGAATCAGAATTCTGTTTTAAAAATTCAATTTTTATGTTGTTTGAGTGCTGTGTATGCTTT CTGTATTTTTATTGTGTCTGTGTTTAGAATCAGAAAGGGTGTTTGAATCTGTTTTAAAAATTC TTTAATTTTCTGTATTTTGTGCTATTTGGTTTATTGGGTATCTAAAAAAGTTGGATCTTGATTT AAAGGCTTTTTAATTTTGTTCTAAAAGTTAAATCTTGATTTTGTTTCAACCTGGTGTGTGCCTTAGGTT TCTGTATTTATGGACTTGGAAACCTTTTAATGATCTAAAAAAGATTGTATTTTAGTTTTATGAGCTG AAAAACCATACAAGTATGATCTGAAAAGTTGAGTCTTGATTTTGACTCAACTTCTGTATGTGT CAATTTCGCATACACCCTCTCCGTACTCCACTTGTGGGATCACTTGCACAATCTATATTGTTGTT ATTTTTGACCTGTGTTTTCTGTTTGTTTGTTTATGATATAAAGAACCTTCCTATTTCTATT TCAATTATTTTGTTTTGAAAAAATTGGAAGCCTTCTTATTGGATGATGATTATGAATGGTGTTCT GTTTTGATTTGTGATATGAAATATCAACTTTAATCTGTCATCTCGTTTTATAGAGTCATGTCTGT TTCGAGTATGTGTGCTTATTTTGTTGAATTTTGTGTACAGATGCAGATATTTGTTAAAACCCTCACTGGA AAGACTATCACCCTTGAGGTGGAAAGCTCAGACACCATTGACAATGTTAAAGCCAAGATCCAGGACAAGG AAGGCATTCCCCCAGACCAGCAGAGGCTGATCTTTGCAGGAAAGCAGCTTGAAGACGGTCGTACTCTAGC TGATTACAACATTCAGAAAGAGTCAACTCTCCACTTGGTGCTCCGTCTTCGTGGTGGC

>SL2.10sc05925 (SL2.10sc.fasta) at 4247068 - 4247228 Promoter = 2000 TGTATCCTAAAATCGAGTTGTGTGATTCGAACTTACTATTGGTAGAATAATTATTTTGATTAACCATCAG ATGAAGTTGCCCCTTTCATAACACATATCACGATTAAGTACGAAAAATATTTCCTAACATAATTT AGTCGAATTCTAACGAATATCACACCTAAATAGATCTCACGTGATAGAAATTTGACTTAGTCAGACTCCA AATATTGTTGTTGGATTATTATATAGGAAGTTGCCATCCAAACGGCACCTAAATTTTCTCCACCGAT CAAAGAAAAGGAGACGTGTCATACTATTATTGGTTACTTCATTGGCAATTTCTTCACAAATTGGCCATTA TATTTGTTTTGTTGTCTGTGTTTTAGAATCAGAATTCTGTTTTAAAAATTCAATTTTTATGTT GTGCTGTGTATGCTTTAATTTTCTGTATTTTTTATTGTGTCTGTGTTTTAGAATCAGAAAGGGTG TCTGTTTTAAAAATTCAATGTTTATGTTGTTTTAGTGTTCTGTATGCTTTAATTTTCTGTATTT TATTTGTTTATTGTGTCTGTGTTTGGAATCAGAAAGGGTGTTTGAATCTGTTTTAAAAATTCAATCTTTA TGTTGTTTGAGTGTCTGTATGATTTAATTTTCTGTATTTTGTGCTATTTGGGTTTATTGGGTATCTAAAAA AGTTGGATCTTGATTTTGTTTTCAATATTCTGTATGCTTTAGTTTTCTGTTAATTGTGTTATTT AACCTGGTGTGTGCCTTAGGTTTCTGTATTTATGGACTTGGAAACCTTTTAATGATCTAAAAAGA TTTTAGTTTTATGAGCTGAAAAACCATACAAGTATGATCTGAAAAGTTGAGTCTTGATTTTGACTC

Comparison of *ubiquitin* genes from other plants

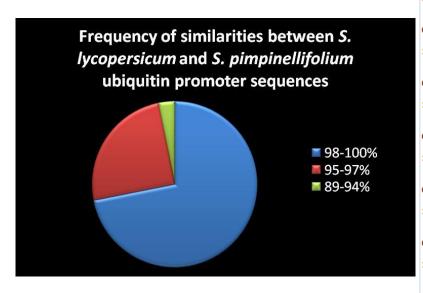


An additional analysis of a draft genome of *S. pimpinellifolium* using the same methodology revealed 32 *ubiquitin* genes. A previous study in soybean showed 25 *ubiquitin* genes (Hernandez-Garcia et al. 2010b). These results confirm that our methodology is robust and likely extracted most of the *ubiquitin* genes present in the genome of tomato.

Homology between ubiquitin promoters from S. lycopersicum and S. pimpinellifolium

- ☐ Similarities/differences between these two promoter groups would give insights into differential gene expression regulation
- ☐ The identified ubiquitin promoters from *S. lycopersicum* were Blasted against the ubiquitin promoters of *S. pimpinellifolium*

Sequence similarities of promoters from both species ranged from 100% to minor bp substitutions, including short INDELs



Query:	855	gctagagttgcggttgagttgaagacgatggcacgtttgctgcactccacaaataacaaa	914
Sbjct:	181	gctagatttacggttgagttgaagacgtcagtacgtttgctacagtccacaaataacaaa	240
Query:	915	gaagaaacataaaagtagggggtcagtacaaacacaggtactgagtaggtatcatcggc	974
Sbjct:	241	gaaagaaacatacaactaggggtt-agtacaaacacaagtactgagtagatatcatcggc	299
Query:	975	taactcaaaatagaaaacagtatatatcagataacatcataaaatcaactaaaatactca	1034
Sbjct:	300	caactcaaaatagaaaacagtatatatcagataacatcataaaatcagctaatatcctta	359
		acatgcagcattttcaattaccataacccttggtcataacaccaagctcatcaacgagga	
Sbjct:	360	gcatgcagcatttaccattaccatcacccttggtcacaacaccaagcacatcaatgagga	419
		ctcacgcctcctcatcatactcatttgggaattaggttcattagattgaatattaaca	
Sbjct:	420	ctcacgcctcctcatcatactcatttgggaactaggttcattaaattgagtatattaaca	479
Query:	1155	tctttcaagattcatttctttattcctctcatgtcggtacgtgacactccgctcctcaa	1214
Sbjct:	480	tctctcaagattcattatctttattcctctcatgttggtacgtgacactctgctcctc-a	538
Query:	1215	tatactatcctcgtgtcagaacgtgacactctgatcctcattctatcctggtgtcgaaat	1274
Sbjct:	539	tatactattctggtgtcggaacgtgacactccgatcctcattctatcctggtgtcggaac	598
Query:	1275	gtgacacccgatccatattctatcatggtaccggaacgtggcacccgatctatatactat	1334
Sbjct:	599	gtgacactcgatccatattctatcttggtaccggaacatggcacccaatccatatactat	658
Query:	1335	cctggtgtcgaaacgtgacactccgatcctcattctatcctggtgtcggaacgtgaca	1392
Sbjct:	659	cctggtgtcggaaggtaacactccgatcctcatatattatcctggtgtcggaacgtgaca	718
Query:	1393	<pre>c-ccgatccatattctatcctggtaccggaatgtggcacctgatccgtatactatcct</pre>	1449
Sbjct:	719	ctcggatcctcatatactatcctggtactggaacgtggcacccgatccatatattatcct	778
Query:	1450	ggtgtcggaacgtgacac 1467	

Sbjct: 779 ggtgttggaacgtgacac 796

Conclusions and biological interpretations

Our methodology is robust to identify promoter sequences of interest in genome sequences ubiquitin genes and their promoters are very conserved among plant species ☐ Identified promoters can be used for further analysis of natural variation, or be cloned and their expression characterized for potential application in the development of GMOs. A complete tutorial on rapid characterization of plant promoters is freely accessible at http://www.jove.com/index/details.stp?id=1733 (Hernandez-Garcia et al. 2010a)

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External Links

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